CM 22617 - SOYABEAN CASEIN DIGEST AGAR PLATE W/ b - LACTAMASE MIXTURE (g - irradiated) (Triple Pack)

INTENDED USE

For cultivation of wide variety of organisms and for inactivation of penicillins, cephalosporins of first, second,third and fourth generation and penems.

PRODUCT SUMMARY AND EXPLANATION

SoyabeanCaseinDigest Agarwith beta-lactamase is used in plates for the detection and enumeration of microorganisms present on surfaces of sanitary importance and also in environmental monitoring of clean room for facilities where production of β-lactam group of antibiotics is carried out.

The media are gamma irradiated in the packaging material to assure a reduction of the microbial load potentially present in the medium, on the dishes, and on the packaging materials. Gamma- irradiation of the product is indicated by an orange to red color of the irradiation indicator stripe on the inner label.

COMPOSITION

Ingredients	Gms / Ltr
Casein enzyme hydrolysate	15.000
Agar	15.000
Papaic digest of Soybean	5.000
Sodium chloride	5.000
Beta-lactamase mixture	500.000 IU

PRINCIPLE

Medium contains Casein enzymic hydrolysate and papaic digest of soybean meal which provide nitrogenous compounds and other nutrients essential for microbial replication. Sodium chloride is added to maintain cellular osmotic equilibrium. Addition of beta-lactamase mixture enables the growth of resistance strains present in the environment of clean roomby inactivating the beta-lactam antibiotics. Agar is used as a solidifying agent.

INSTRUCTION FOR USE

Eitherstreak,inoculate orsurface spread the test inoculum aseptically on the plate. Alternatively, these plates can also be used as settle plates forenvironmental monitoring..

QUALITY CONTROL SPECIFICATIONS

Appearance	:	Light amber color, clear to slightly opalescent gel.
Quantity of Medium	:	18 ml of medium in 55 mm plates.
pH (at 25°C)	:	7.3± 0.2
Dose of irradiation:	:	15-25 kGy
Sterility Check	:	Passes release criteria

INTERPRETATION

Cultural characteristics observed after incubation .

Growth Promotion Test

Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Recovery	Incubation Temperature	Incubation Period
Bacillus subtilis	6633	50-100	Luxuriant	>=70%	30-35°C 30-	18-24 hours
Staphylococcus aureus	25923	50-100	Luxuriant	>=70%	35°C 30-35°C	18-24 hours
Escherichia coli	25922	50-100	Luxuriant	>=70%	30-35°C 30-	18-24 hours
Pseudomonas aeruginosa	27853	50-100	Luxuriant	>=70%	35°C 30-35°C	18-24 hours
Streptococcus pneumonia	6305	50-100	Luxuriant	>=70%	30-35°C 20-25	18-24 hours
Salmonella typhimurium	14028	50-100	Luxuriant	>=70%	℃ 20-25 ℃	18-24 hours
Enterococcus faecalis	29212	50-100	Luxuriant	>=70%		18-24 hours
Candida albicans	10231	50-100	Luxuriant	>=70%		<=5 days
Aspergillus brasiliensis	16404	50-100	Luxuriant	>=70%		<=5 days

Cultural Response

Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Recovery	Incubation Temperature	Incubation Period
Escherichia coli	25922					
w/o antibiotic			Luxuriant	>=70%	30-35°C	18-24 hours
w/ Cephalothin			Luxuriant	>=70%	30-35°C	18-24 hours
w/ Cefotaxime			Luxuriant	>=70%	30-35°C	18-24 hours
w/ Ceftazidime		50-100	Luxuriant	>=70%	30-35°C	18-24 hours
w/ Imipenem			Luxuriant	>=70%	30-35°C	18-24 hours
w/ Ertapenem			Luxuriant	>=70%	30-35°C	18-24 hours
w/ Meropenem			Luxuriant	>=70%	30-35°C	18-24 hours
Staphylococcus aureus				>=70%	30-35°C	18-24 hours
w/o antibiotic	25923					
w/Penicillin			Luxuriant	>=70%	30-35°C	18-24 hours
w/ Cephalothin			Luxuriant	>=70%	30-35°C	18-24 hours
w/ Cefotaxime			Luxuriant	>=70%	30-35°C	18-24 hours
w/ Ceftazidime		50-100	Luxuriant	>=70%	30-35°C	18-24 hours
w/ Imipenem			Luxuriant	>=70%	30-35°C	18-24 hours
w/ Ertapenem			Luxuriant	>=70%	30-35°C	18-24 hours
w/ Meropenem			Luxuriant	>=70%	30-35°C	18-24 hours
			Luxuriant	>=70%	30-35°C	18-24 hours

PACKAGING:

Triplelayeredpacking containing 5 No. of plates with one silica gel desiccant bag packed inside it.

STORAGE

Onreceipt, store the plates at 15–30 °C. Avoid freezing and overheating. Do not open until ready to use. Prepared plates stored in their original sleeve wrapping until just prior to use may be inoculated up to the expiration date and incubated for recommended incubation times. Allow the medium to warm to room temperature before inoculation. Product Deterioration: Do not use plates if they show evidence of microbial contamination, discoloration, drying, cracking or other signs of deterioration.

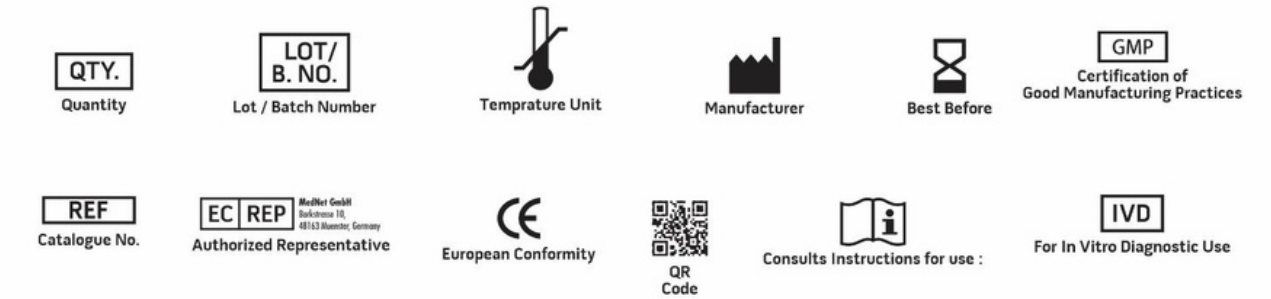
DISPOSAL

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

REFERENCES

1. HallandHartnett, 1964, Public Hlth. Rep., 79:1021.
2. Richardson (Ed)., 1985, Standard Methods for the Examination of Dairy Products, 15th ed., APHA, Washington, D.C.
3. MacFaddin J.F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. I, Williams and Wilkins, Baltimore.
4. Brummer, 1976, Appl. Environ. Microbiol., 32:80.
5. Favero (Chairm), 1967, Biological Contamination Control Committee, a state of the art report.,Am. Assoc. for contamination control.

Murray PR, Baron, Pfaller, and Yolken (Eds.), 2003, In Manual of Clinical Microbiology, 8th ed., ASM, Washington, D.C.



NOTE: Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices.

*For LabUse Only

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