CM 22620 - SOYABEAN CASEIN DIGEST AGAR PLATE W/LECITHIN AND POLYSORBATE 80 (TRYPTONE SOYA AGAR PLATE W/LECITHIN AND POLYSORBATE 80 (γ-IRRADIATED) (TRIPLE PACK)

INTENDED USE

For determining efficiency of sanitization of containers, equipments, surfaces, water miscible cosmetics etc.

PRODUCT SUMMARY AND EXPLANATION

Soyabean casein Digest Agar Plate w/ Lecithin and Polysorbate 80 is RODAC (Replicate Organism Detection and Counting) plates are recommended for theisolation of microorganisms from environmental surfaces and is used primarily to monitor microbial contamination,enumerate the number of microbial colonies growing on a variety of surfaces sanitized with quaternary ammoniumcompounds, and to assist in determining surface sanitation.

The media are gamma irradiated in the packaging material to assure a reduction of the microbial load potentially present in the medium, on the dishes, and on the packaging materials. Gamma- irradiation of the product is indicated by an orange to red color of the irradiation indicator stripe on the inner label.

COMPOSITION

Ingredients	Gms / Ltr
Casein enzymatic hydrolysate	15.000
Agar	15.000
Papaic digest of Soybean meal	5.000
Polysorbate 80 (Tween 80)	5.000
Sodium chloride	5.000
Lecithin	0.700

PRINCIPLE

Medium contains Casein enzymatic hydrolysate and papaic digest of soyabean meal which helps to provide nitrogenous compounds and other nutrients essential for microbial replication. Sodium chloride is added to maintain cellular osmotic equilibrium. Lecithin and polysorbate 80 are added to the formulation to neutralize germicidal or disinfectant residues. Neutralization of these residues reduces their inhibitory effect which ultimately results in lowering of microbial count. Quaternary ammonia compounds are neutralized by lecithin, while phenolic disinfectants and hexachlorophene are neutralized by polysorbate 80. Together, lecithin and polysorbate 80 neutralize ethanol.

INSTRUCTION FOR USE

Eitherstreak,inoculateorsurface spread the test inoculum aseptically on the plate. Alternatively, these plates can also be used as contact platesfor environmental monitoring.

QUALITY CONTROL SPECIFICATIONS

Appearance	:	Light amber color, clear to slightly opalescent gel.
Quantity of Medium	:	15-18 ml of medium in 55 mm plates.
pH (at 25°C)	:	7.3± 0.2
Dose of irradiation:	:	15-25 kGy
Sterility Check	:	Passes release criteria

INTERPRETATION

Cultural characteristics observed after inoculation of 50-100 CFU, on incubation at 30- 35 °C for 18 – 24 hours for bacteria and at 30- 35 °C and 20-25°C for ≤ 5 days for fungus. Recovery rate is considered 100% for bacteria growth on Soya Agar and fungus growth on Sabouraud Dextrose Agar.

Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Recovery	Incubation Temperature	Incubation Period
Bacillus subtilis	6633	50-100	Luxuriant	>=70%	30-35°C	18-24 hours
Staphylococcus aureus	6538	50-100	Luxuriant	>=70%	30-35 °C	18-24 hours
Escherichia coli	8739	50-100	Luxuriant	>=70%	30-35 °C	18-24 hours
Pseudomonas aeruginosa	9027	50-100	Luxuriant	>=70%	30-35 °C	18-24 hours
Salmonella typhimurium	14028	50-100	Luxuriant	>=70%	30-35 °C	18-24 hours
Candida albicans	10231	50-100	Luxuriant	>=70%	30-35 °C	24-48 hours
Candida albicans	10231	50-100	Luxuriant	>=70%	20-25 °C	48-72 hours
Aspergillus brasiliensis	16404	50-100	Luxuriant	>=70%	30-35 °C	48-72 hours
Aspergillus brasiliensis	16404	50-100	Luxuriant	>=70%	20-25°C	72-120 hours

PACKAGING:

Triplelayeredpacking containing 5 No. of plates with one silica gel desiccant bag packed inside it.

STORAGE

Onreceipt, store the plates at 15–30 °C. Avoid freezing and overheating. Do not open until ready to use. Prepared plates stored in their original sleeve wrapping until just prior to use may be inoculated up to the expiration date and incubated for recommended incubation times. Allow the medium to warm to room temperature before inoculation. Product Deterioration: Do not use plates if they show evidence of microbial contamination, discoloration, drying, cracking or other signs of deterioration.

DISPOSAL

Afteruse,prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

REFERENCES

1. The United States Pharmacopoeia. 2009. Amended Chapters 61, 62 & 111, The United States Pharmacopoeial Convention Inc., Rockville, MD.
2. Hall and Hartnett, 1964, Public Hlth. Rep., 79:1021.
3. Richardson (Ed)., 1985, Standard Methods for the Examination of Dairy Products, 15th ed., APHA, Washington, D.C.
4. MacFaddin J.F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. I, Williams and Wilkins, Baltimore.
5. Brummer, 1976, Appl. Environ. Microbiol., 32:80.
6. Erlandson A.L. Jr and Lawrence C.A. 1953, Inactivating medium for hexachlorophene (G-11) types of compounds and some substituted phenolic disinfectants, Science, 118, 274-276.
7. Favero (Chairm), 1967, Biological Contamination Control Committee, a state of the art report., Am. Assoc. for contamination control

NOTE: Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices.

*For LabUse Only