

CM 21,338 – ORANGE SERUM AGAR

INTENDED USE

For cultivation and enumeration of microorganisms associated with the spoilage of citrus products, cultivation of Lactobacilli, other aciduric organisms and pathogenic fungi.

PRODUCT SUMMARY AND EXPLANATION

Fruit juices are generally acidic, with pH values ranging from approximately 2.4 for lemon juice, to 4.2 for tomato juice. The low pH of these foods is selective for yeast, moulds and a few groups of aciduric bacteria. The microorganisms of greatest significance in citrus juices are the lactic acid bacteria, primarily species of Lactobacillus and Leuconostoc, yeast and moulds. Microbial spoilage of these citrus fruit juices is most commonly due to aciduric microbes such as lactic acid bacteria and yeast. The lactic acid bacteria include Lactobacillus fermentum, L. plantarum, and Leuconostoc mesenteroides. Orange Serum Agar is recommended by APHA for cultivation of Lactobacilli and other aciduric organisms. Orange Serum Agar was originally developed by Murdock et al and Hays for examining citrus concentrates. Hays and Reister further used this medium for studying the spoilage of orange juice. Dehydrated agar medium containing orange serum was reported by Stevens. Orange Serum Broth is used to initiate growth of saprophytic, pathogenic fungi in small samples.

COMPOSITION

Ingredients	Gms / Ltr
Tryptone	10.000
Yeast extract	3.000
Dextrose (Glucose)	4.000
Dipotassium hydrogen phosphate	2.500
Orange serum	9.000
Agar	17.000

PRINCIPLE

The medium consists of Tryptone which provides essential nitrogenous, carbonaceous compounds, long chain amino acids and other essential nutrients. Dextrose (Glucose) serves as the fermentable carbohydrate and energy source. Yeast extract supplies B-complex vitamins, which stimulate growth. Orange serum provides an optimal environment for the recovery of acid tolerant microorganisms from citrus fruit products.

INSTRUCTION FOR USE

- Dissolve 45.5 grams in 1000 ml purified / distilled water.
- Heat to boiling to dissolve the medium completely.
- Sterilize by autoclaving at 15 psi pressure (121°C) for 15 minutes. AVOID OVERHEATING.
- Cool to 45-50°C. Mix well and pour into sterile Petri plates.

QUALITY CONTROL SPECIFICATIONS



Appearance of Powder : Cream to yellow homogeneous free flowing powder.
 Appearance of prepared medium : Light amber coloured clear solution in tubes.
 pH (at 25°C) : 5.5 ± 0.2

INTERPRETATION

Cultural characteristics observed after incubation.

Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Recovery	Incubation Temperature	Incubation Period
Candida albicans	10231	10-100	Good-luxuriant	≥50%	25-30°C	40-48 Hours
Lactobacillus acidophilus	4356	50-100	Good-luxuriant	≥50%	35-37°C	40-48 Hours
Lactobacillus fermentum	9338	50-100	Good-luxuriant	≥50%	35-37°C	40-48 Hours
Leuconostoc mesenteroides	12291	50-100	Good-luxuriant	≥50%	35-37°C	40-48 Hours
Saccharomyces cerevisiae	9763	10-100	Good-luxuriant	≥50%	25-30°C	40-48 Hours
Aspergillus niger	16404	10-100	Good-luxuriant	≥50%	25-30°C	40-48 Hours

PACKAGING:

In pack size of 500 gm bottles.

STORAGE

Dehydrated powder, hygroscopic in nature, store in a dry place, in tightly-sealed containers between 10-25°C and protect from direct sunlight. Under optimal conditions, the medium has a shelf life of 4 years. When the container is opened for the first time, note the time and date on the label space provided on the container. After the desired amount of medium has been taken out replace the cap tightly to protect from hydration.

Product Deterioration: Do not use if they show evidence of microbial contamination, discoloration, drying or any other signs of deterioration.

DISPOSAL

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.



REFERENCES

1. Hays G. L., 1951, Proc. Florida State Hort. Soc., 54:135.
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3. Hays G. L. and Reister D. W., 1952, Food Technol., 6:186.
4. orgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
5. MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. 1, Williams and Wilkins, Baltimore.
6. Murdock P. I., Folinazzo J. F., and Troy V. S., 1951, Food Technol., 6:181.
7. Salfinger Y., and Tortorello M.L. Fifth (Ed.), 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.
8. Stevens J. W., 1954, Food Technol., 8:88.



NOTE: Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices.
*For Lab Use Only

